1	SJST MANUSCRIPT TEMPLATE FOR A TEXT FILE
2	
3	Original Article
4	Effect of supply and demand of phloem sugar on the proportion of brachypterous
5	forms of the rice pest Nilaparvata lugens Stål (Hemiptera: Delphacidae).
6	
7	Piyanut Nasonthron ¹ , Anoma Dongsansuk ² , Ubon Tangkawanit ^{1*}
8	
9	¹ Department of Entomology and Plant Pathology, Faculty of Agriculture, Khon Kaen
10	University, Khon Kaen, 40002, Thailand
11	² Department of Agronomy, Faculty of Agriculture, Khon Kaen University, Khon Kaen,
12	40002, Thailand
13	* Corresponding author, Ubon Tangkawanit
14	Email address: ubonta@kku.ac.th
15	
16	Abstract
17	The sugar supplied by rice and the demand by N. lugens can affect the
18	proportion of brachypterous forms in a population. Experiments were conducted to
19	estimate the food ingestion by N. lugens per capita, to determine total food demand in a
20	rice field, and to find the relation of food supply to the proportion of the brachypterous
21	form. The results revealed that the sucrose content tended to decrease with the age of
22	rice. However, the food demand by the N. lugens population dramatically increased in
23	40-89-day-old rice. The proportion of sugar supply to insect demand decreased as the
24	insect population increased. The relative abundance of short-wing form insects

increased when the ratio of sugar supply to insect demand was ≥ 0.02 , but was decreased when the ratio was lower than 0.01. These results are useful for predicting *N. lugens* dispersal related to sugar supply in rice fields.

28

29 Keywords: brown planthopper, food demand, food ingested, macropterous, sucrose.

30

31 **1. Introduction**

Rice is one of the three most important grain crops in the world (Chauhan, 32 33 Jabran, & Mahajan, 2017). It is a staple food for much of the world's population, with a production estimate of 519.5 million tons from 2022-2023 (Food and Agriculture 34 Organization of the United Nations [FAO], 2023). Important factors that are 35 contributing to the declining profitability in rice production are insect pests and 36 diseases. Nilaparvata lugens Stål (Hemiptera: Delphacidae) is an important pest of rice 37 38 cultivation. It has serious effects on the growth of rice plants, resulting in very large 39 yield losses in rice-growing areas (Jena et al., 2018). The damage is directly related to the desiccation of rice plants, which occurs as a result of insects consuming the plant 40 41 fluids (referred to as 'hopper burn'), or is indirectly related to the transmission of viral diseases called 'rice grassy stunt' and 'ragged stunt virus' (Rivera, Ou, & Iida, 1966). 42 43 Outbreaks of N. lugens often result in high economic losses. For example, in 2005, 1,880,000 tons of rice was lost due to N. lugens damage in China (Hu et al. 2014). 44

In the adult stage, there are two winged forms of adult *N. lugens*, a macropterous (long-winged) form and a brachypterous (short-winged) form (Figure 1). The brachypterous form has high fecundity and is flightless. This form is dominant if the insects have access to plenty of host plants. In contrast, the macropterous form has low fecundity (Xayyasin, Khlibsuwan, & Tangkawanit, 2014) and its population increases in older rice. The long wing of macropterous forms helps their dispersal. To predict *N. lugens* population dynamics, knowledge of the abiotic and biotic factors, such as accumulated degree days, fecundity, survivorship, and functional and numerical response of natural enemies and weather data, that influence their populations is important. Additionally, the changing balance of wing dimorphism types is important for estimating the timing of local insect dispersal in rice cultivation.

Wing dimorphism is considered to be influenced by environmental factors, 56 57 population density, nutrition, juvenile hormones, interspecific interactions and abiotic factors such as photoperiod (Zhi-Fang, Ju-Long, Juan, Chao, & Xiang-Dong, 2014). 58 The most important influence on the wing dimorphism of *N*. *lugens* is host plant quality 59 (Kisimoto, 1956; Iwanaga, Tojo, & Nagata, 1985; Xu et al., 2015; Liu et al., 2020). 60 Romadhon, Koesmaryono, and Hidayati (2017) found that the population of N. lugens 61 62 becomes macropterous and emigrates during the 4-5 weeks after transplantation. Syobu, Mikuriya, Yamaguchi, Matsuzaki, and Matsumura (2002) reported that the 63 incidence of brachypterous females dramatically decreased approximately 75-85 days 64 65 after the rice was transplanted. A possible reason for this might be that the nutritional conditions tend to decline in older rice (Baqui & Kershaw, 1993; Wu, Yu, Tao, & Ren, 66 67 1994; Xayyasin et al., 2014).

Nilaparvata lugens feeds on rice phloem sap, which contains large amounts of
sugars (Kikuta, Kikawada, Hagiwara-Komoda, Nakashima, & Noda, 2010). Okamura,
Hashidaa, Hirosea, Ohsugia, and Aokia (2016) found that sucrose was the main soluble
sugar in rice stems, but glucose and fructose were also present. Deepa, Pillai, and
Murugesan (2016) revealed that the total sugar content was found to differ significantly

in rice of different ages and varieties. Lin, Xu, Jiang, Lavine, and Lavine (2018) found 73 that the glucose concentration in older rice plant is much greater than in younger ones. 74 Knowing the quantity of food ingested by N. lugens per capita is important for 75 estimating the food demand of the insects in a paddy field. We hypothesized that the 76 77 supply of sugar from rice and the demand for it by insects affect the proportion of the brachypterous form of N. lugens. If the sugar supply is lower than the insect demand, 78 79 the brachypterous form will decrease. The objectives of this research were: - (1) to estimate the food ingestion by N. lugens per capita for both nymphs and adults; (2) to 80 estimate the insect food demand in rice field conditions; and (3) to determine the ratio 81 82 of food supply to the proportion of the brachypterous form of *N. lugens*.

83

84 2. Materials and Methods

85 **2.1 Insect rearing**

86 *Nilaparvata lugens* were collected from a paddy field and released in a cage

 $(50\times70\times100 \text{ cm})$ made of a wooden frame with a wire mesh covering the top and

sidewalls, maintained at the Department of Entomology and Plant Pathology, Faculty of

- Agriculture, Khon Kaen University. Twenty 40-day-old rice plants of the variety
- 90 Taichung native 1 (TN1) were placed in the cage as the host plant for feeding and
- oviposition. Old rice plants were replaced with new plants after 10 days.
- 92 2.2 Food demand

93 2.2.1 Assimilation and ingestion of food

94 The experiment was conducted in a 7 cm diameter plastic pot containing a 45-day-old

95 rice plant (TN1). Nymphs (third nymphal stage) and adults of *N. lugens* were starved

96	for 2 hours before use. Each individual was weighed and released in a parafilm sachet
97	(5x10 cm), which was attached to the base of the plant (3 cm above the soil surface).
98	There were 20 replicates for both nymphs and adults. After 24 hours, the insect was
99	removed from the sachet and weighed separately. The honeydew on the parafilm
100	sachets was also weighed. The sachet and honeydew were weighed together, then the
101	honeydew was removed and the sachet was reweighed. A control was conducted to
102	assess the loss of body weight from catabolism, with moist cotton being provided
103	instead of a rice plant.
104	Food assimilation was calculated by the method of Smith, Khan, and Pathak
105	(1994).
106	Food assimilated = W1 x $[(C1-C2)/C1] + (W2-W1)$, where
107	W1 = Initial weight of the insect,
108	W2 = Final weight of the insect,
109	C1 = Initial weight of the control insect,
105	
110	C2 = Final weight of the control insect
111	
112	Food ingested = Food assimilated + weight of the honeydew.
113	Food assimilated, honeydew excreted and food ingested by nymph and adult <i>N</i> .
114	lugens on the Taichung native 1 (TN1) rice variety were compared by a paired t-test
115	(p=0.05) using Statistix10.
116	

117 2.2.2 Field experiment

An insect outbreak was simulated and studied in the field research area of the 118 119 Department of Entomology and Plant Pathology, Faculty of Agriculture, Khon Kaen 120 University, using Jasmine rice (KDML 105 variety), a susceptible variety that is the 121 preferred variety cultivated in Thailand and other Asian countries. Forty-day-old seedlings of KDML 105 were transplanted to a field with an area of 400 m². Four field 122 123 cages made of iron frames (1.25x1.25x2 m), covered with a fine mesh on the tops and 124 sidewalls with a door with a zipper-opening on one side, were positioned to enclose the 125 rice plants. There were 25 rice hills per cage at a spacing of 25x25 cm. Three adults of 126 *N. lugens* per rice hill were released (75 adults per cage with 2:1 female to male ratio). Nymphs and adults (brachypterous and macropterous) of *N. lugens* were recorded by 127 direct counting on 13 hills per cage every week. One rice plant per hill was collected for 128 129 further sugar analysis.

Food demand was estimated by multiplying the food ingested by the
populations of nymphs and adults (total food demand = food ingested x insect
population).

133 **2.3 Sugar analysis**

134 2.3.1 Rice plant material

Pieces of rice stem (5 cm long and cut 3 cm above the soil surface) were
collected from the experimental cages. A total of 0.1 grams of each rice stem was cut
into smaller pieces placed in a test tube, and 3 ml of 80% ethanol was added, held in a
boiling water bath at 100 °C for 1 min and then transferred to a 65 °C water bath. After
1 hour, the supernatant was transferred to a new tube. Residual solid rice tissues were

140	extracted with 3 ml of 80% ethanol and warmed in a 65 °C water bath for 1 hour two			
141	additional times. The supernatants of the three extractions were combined for the			
142	determination of sucrose content.			
143	2.3.2 Sucrose content			
144	Sucrose content was analyzed in supernatants from the plant extraction using the			
145	method described by Robbins and Pharr (1987). A total of 500 μ l of the supernatant was			
146	transferred to a tube, and then 0.25 ml of 1% resorcinol in 95% ethanol and 0.75 ml of			
147	30% HCL were added. The solution was incubated in an 80 °C water bath for 10 min.			
148	The tube was removed and cooled to room temperature. Absorbance at wavelength 520			
149	nm was measured with a spectrophotometer. Sucrose content was quantitated by			
150	comparison to sucrose standards.			
151	2.4 Sugar supply to insect demand			
151 152	2.4 Sugar supply to insect demand The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was			
152	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was			
152 153	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was determined as			
152 153 154	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was determined as S2D = CH2O/Food demand.			
152 153 154 155	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was determined as S2D = CH2O/Food demand. The data for food demand were calculated by the food ingestion of nymphs and			
152 153 154 155 156	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was determined as S2D = CH2O/Food demand. The data for food demand were calculated by the food ingestion of nymphs and adults multiplied by the number of insects examined in the cage for each observation.			
152 153 154 155 156 157	The sugar supply from rice to <i>N. lugens</i> demand (S2D) for each observation was determined as S2D = CH2O/Food demand. The data for food demand were calculated by the food ingestion of nymphs and adults multiplied by the number of insects examined in the cage for each observation. CH2O was the sugar content in the rice in each observation.			

161 relative abundance = Nb/(Nb+Nm),

where Nb = number of brachypterous forms, and Nm = number of macropterousforms.

164

165 **3. Results**

166 **3.1 Food demand**

167 **3.1.1** Assimilation and ingestion of food

The amount of food assimilated, honeydew excreted and food ingested by
nymphs and adult *N. lugens* on 45-day-old TN1 rice after 24 hours are shown in Table
1. The quantities of those 3 parameters were significantly higher in the adult stage than
in the nymphal stage. From this experiment, the food ingestion of *N. lugens* per capita
for nymphs and adults was 1.21 and 10.60 mg/day, respectively.

173 **3.1.2 Food demand in rice field conditions**

The population of *N. lugens* was recorded every week. Food demand for each 174 175 week was estimated as shown in Table 2. The results revealed that the number of nymphs slightly increased on rice that was 54-82 days old. The first released adult 176 population slightly decreased during the first to third weeks. When the rice age was 61 177 days, the first released adult population died. The total number of adults was 0 in 61-day 178 old rice. Then, the 2nd adult population increased and developed to adult in the rice field. 179 180 The total food demand increased during vegetative growth until the rice age was 89 days old (1,033.97 mg/hill). After the rice was in reproductive growth, the insect 181 population and food demand both dropped. 182

184 **3.2 Rice sucrose content**

The sucrose content of rice infested by *N. lugens* was significantly lower than that of non-infested rice at 47–103 days old. The sucrose content of infested rice and non-infested rice was dramatically decreased at 40–61 days. After 61 days, the sucrose content of the non-infested rice was stable, whereas the sucrose content of the infested rice had decreased slightly (Figure 2).

191 **3.3** Sugar supply to insect demand and relative abundance

The proportions of sugar content and insect demand for each age of the rice plants are presented in Table 3. The results indicated that the ratio of sugar supply to insect demand tended to increase when the rice was 40–54 days old. Thereafter, the ratio decreased dramatically at 54–68 days old. Sugar supply to insect demand was lowest at 89 days old.

197 The relative abundance of the short-wing form in the first release was 0.5, and 198 then the value subsequently increased until the rice was 54 days old. When the rice was 199 61 days old, the adults of the first released population had died and were no longer found in the experimental cage. For this rice age, the ratio of sugar supply to insect 200 demand is not zero (0.09) because there are some nymphs of 2^{nd} population from eggs 201 laid by the first generation in the field. After that, the 2nd adult population of 202 203 brachypterous adults, arising, began to increase until the rice was 75 days old. After the 204 rice was 82 days old, the supply to demand ratio trended to lower than 0.01, and the 205 relative abundance value was lower than 0.6. The results indicate that there were fewer 206 brachypterous forms than macropterous forms. The relative abundance of short-wing form then slightly decreased to zero when the rice was 103 days old. 207

208

209 **4. Discussion**

The quantity of food ingested was higher in the adult stage than in the nymphal stage. This result is similar to that obtained for nymphs and adults of the grasshopper *Oxya hyla hyla*, which is a common rice pest (Ghosh, Haldar, & Mandal, 2014). This result may be related to the larger size of the adults, which requires them to ingest more 214 food, nutrients and energy. In addition, adults expend most of the energy from their food in reproduction (Ghosh et al. 2014). The calculated values of food ingested found 215 216 in this experiment differed from those from previous studies (Senthil-Nathan, Choi, Paik, Seo, & Kalavani, 2009; Latif et al., 2012; Mollah, Samad, Hossain, & Khatun, 217 218 2011). There are some ecological factors influencing the parameters used for the calculation of food ingestion, such as rice age and rice variety. Baqui and Kershaw 219 (1993) found that the honeydew secreted by N. lugens was lower on 90 days old rice 220 221 plants than on younger rice plants. Latif et al. (2012) and Mollah et al. (2011) reported 222 that the amount of food ingested and assimilated was significantly decreased when N. lugens was reared on a resistant variety of rice compared to a susceptible variety of rice. 223 224 In addition, Lu et al. (2007) and Wu et al. (2017) revealed that ingestion rates of the brown planthopper were increased when feeding on N-fertilized plants. Food ingested 225 in this experiment was estimated in a rice variety (TN1) susceptible to N. lugens. 226 227 Therefore, the results indicated the amount of food ingested when food is plentiful and 228 favoured by insect pests. Forty-day-old rice was studied because this age of rice has 229 often been detected in N. lugens outbreaks.

230 The food demand of N. lugens in the rice field depended on the size of the insect population. The highest population number of N. lugens was usually found when 231 232 rice was in the late vegetative stage. Sawada, Subroto, Suwardiwijaya, Mustaghfirin, and Kusmayadi (1992) and Khlibsuwan, Hanboonsong, Pannangpetch and Sriratanasak 233 234 (2014) showed that there were approximately 3 generations of N. lugens in rice fields, 235 which corresponds to our results. In this experiment, the initial population died within 2-3 weeks after release. Then, a 2^{nd} generation emerged from the eggs that had been 236 oviposited by the 1st generation (Table 2). The population increased until the rice was 237

89 days old; then, a final generation of long-winged morphs became established in therice field.

240 Sucrose is the major sugar product of photosynthesis in rice plants and is the main soluble sugar in the rice stem (Kikuta et al., 2010; Okamura et al., 2016). Sucrose 241 242 is transported from source tissues to various sink tissues by the phloem to sustain plant 243 development, such as pollen development and pollen tube growth before the heading 244 stage. In developing seeds, the phloem releases sucrose into maternal tissues to produce 245 the grains (Jung & Im, 2005). Therefore, the sucrose content in the rice tilling of older 246 rice was lower. A decrease in the total sucrose content with increasing plant age was observed in this experiment. This result corresponds with the report of Deepa et al. 247 (2016) that the total sugar content was decreased 20, 40, and 60 days after rice was 248 transplanted. The trend of sucrose content in rice plants was opposite to that of the 249 glucose content reported by Lin et al. (2018). Sucrose is a disaccharide consisting of 250 251 one glucose and one fructose molecule; therefore, it may be that decreasing sucrose 252 content may have resulted in increase of glucose content arising through hydrolysis of sucrose in older plants. 253

The main food source for N. lugens is sugar. It was observed that the sugar 254 255 supply to insect demand decreased with an increasing insect population. The relative 256 abundance of short wing form was highest when the rice age was 75 days. The ratio of 257 the supply by plant and the demand by the insect at this age was 0.02. This amount of 258 supply may be sufficient for the insects. However, after the rice was 82 days old, when the sugar supply to insect demand was lower than 0.01, the brachypterous form was less 259 abundant than the macropterous adults. This is the critical value that indicates the 260 imminent population migration of *N. lugens*. This result is similar to those in a report by 261

262 Syobu et al. (2002) that showed brachypterous females dramatically decrease 263 approximately 75-85 days after rice is transplanted and that rice plant stage affects the female wing-form ratio. At 96 days into the experiment, the rice reached the booting 264 stage of reproductive growth and developed a panicle primordium. The sugar may 265 266 transfer to sink cells for panicle development. Therefore, the food supply was very low and not enough to support a high density of insects. Compared to non-infested plants, 267 the sugar content in infested plants was very low, and most of the sugar was lost 268 269 because of insect sucking. Some rice plants showed symptoms of hopper burn and 270 turned from green to reddish-brown at 89 days (S2D=0) (Figure 3). Therefore, this rice symptom indicates a high density of the macropterous form and that the insect is ready 271 272 for dispersal to the next locality. This is a critical point for vigilance in detecting outbreaks in neighboring areas. However, the amount of sugar content varied depending 273 274 on the rice variety (Deepa et al. 2016) and macropterous forms may be present earlier in low-sugar varieties than in high-sugar varieties. 275

Iwanaga et al. (1985) and Ayoade, Sunao, and Sumio (1996) reported that high 276 277 levels of juvenile hormones in the early nymphal stage induced the brachypterous form. 278 However, juvenile hormones are affected by food quantity. Saxena, Okech, and Liquid (1981) suggested that inadequate food affects juvenile hormones released from the 279 280 corpus allatum. It is possible that a starvation situation brought about by an older rice age may result in a change in the level of juvenile hormones, which affects the wing 281 282 form of *N. lugens*. This report corresponded to the report of Simpson and Raubenheimer 283 (1993), that metabolic activities during insect development are dependent on the quality of food. When foods are nutritionally imbalanced, insect herbivores have to adapt by 284 evolving an appropriate behavioural and physiological mechanism (Behmer, 2009). 285

Zhang, Mao, & Liu (2023) revealed that 4 developmental regulated genes of wing, 286 NIInR1, NIInR2, NIAkt, and NIFoxo were expressed in the short-winged adults, but 287 silenced in the long winged adults. Lin et al. (2018) showed that two insulin receptors 288 (NIInR1 and NIInR2) regulated wing type of N. lugens. Recently, Liu et al. (2020) 289 290 revealed that ultrabithorax (Ubx) is a key regulator for promoting short wing form in N. lugens. They suggested that a high quality of plant nutrition at a later stage of nymph 291 292 increased NIInR2 expression. NIInR2 induced high level of Ubx and consequently 293 suppress the NIInR1 resulting in short wing form in N. lugens. Based on the present 294 results and earlier researcher, food is identified as the important factor for wing development in N. lugens. However, some abiotic factors such as temperature and 295 296 photoperiod may influence wing dimorphism. The results of this study should prove useful for predicting N. lugens dispersal in rice fields. 297

298 **5.** Conclusions

299 Nilaparvata lugens Stal's primary food source is sugar from rice phloem sap. Adults consumed 10.6 mg of food each day, while nymphs only consumed 1.21 mg. As 300 301 rice grew older, the sugar concentration tended to diminish. The number of 302 brachypterous forms in a population can vary depending on the amount of sugar that is produced by rice and the demand from N. lugens. The proportion of sugar supply to 303 304 insect demand decreased with an increasing insect population. The relative abundance of short-wing form insects increased when the sugar supply to insect demand was 305 306 ≥ 0.02 , and became decreased when the sugar supply to insect demand was lower than 307 0.01. This information is useful for N. lugens population prediction in the rice production. 308

309 Acknowledgments

310	This work was financially supported by Khon Kaen University. We would like			
311	to thank Assoc. Prof. Dr. Wirote Khlibsuwan who provided expertise that greatly			
312	assisted the research.			
313				
314	References			
315	Ayoade, O., Sunao, M., & Sumio, T. (1996). Induction of macroptery, precocious			
316	metamorphosis, and retarded ovarian growth by topical application of precocene II,			
317	with evidence for its non-systemic allaticidal effects in the brown planthopper,			
318	Nilaparvata lugens. Journal of Insect Physiology, 42, 529–540. doi:10.1016/0022-			
319	1910(96)00135-7			
320	Baqui, M.A., & Kershaw, W.J.S. (1993). Effect of plant age on host preference,			
321	honeydew production and fecundity of Nilaparvata lugens (stal) (Hom.,			
322	Delphacidae) on rice cultivars. Journal of Applied Entomology, 116, 133–138.			
323	doi:10.1111/j.1439-0418.1993.tb01179.x			
324	Behmer, S.T. (2009). Insect herbivore nutrient regulation. Annual Review of			
325	Entomology, 54: 165–187. doi:10.1146/annurev.ento.54.110807.090537			
326	Chauhan, B.S., Jabran, K., & Mahajan, G. (2017). Rice production Worldwide. Springer			
327	International Publishing AG, Switzerland.			
328	Deepa, K., Pillai, M.A., & Murugesan, N. (2016). Biochemical bases of resistance to			
329	brown planthopper (Nilaparvata lugens) (Stål) in different rice accessions.			
330	Agricultural Science Digest, 36, 102–105. doi:10.18805/asd.v36i2.10627			

331	Food and Agriculture Organization of the United Nations, (2023). World Food				
332	Situation. Retrieved from https://www.fao.org/worldfoodsituation/csdb/en/.				
333	Accessed February 2023.				
334	Ghosh, S., Haldar, P., & Mandal, D.K. (2014). Suitable food plants for mass rearing of				
335	the short-horn grasshopper Oxya hyla hyla (Orthoptera: Acrididae). European				
336	Journal of Entomology, 111, 448-452. doi: 10.14411/eje.2014.038				
337	Hu, G., Fang, L., Bao-Ping, Z., Ming-Hong, L., Wan-Cai, L., Feng, Z., Xiang-Wen, W.,				
338	Gui-Hua, C., & Xiao-Xi, Z. (2014). Outbreaks of the brown planthopper				
339	Nilaparvata lugens (Stål) in the Yangtze River Delta: Immigration or local				
340	reproduction. PLoS One, 9, e88973. doi:10.1371/journal.pone.0088973				
341	Iwanaga, K., Tojo, S., & Nagata, T. (1985). Immigration of the brown				
341 342	Iwanaga, K., Tojo, S., & Nagata, T. (1985). Immigration of the brown planthopper, <i>Nilaparvata lugens</i> , exhibiting various responses to density in				
342	planthopper, Nilaparvata lugens, exhibiting various responses to density in				
342 343	planthopper, <i>Nilaparvata lugens</i> , exhibiting various responses to density in relation to wing morphism. <i>Entomologia Experimentalis et Applicata, 38</i> , 101–				
342 343 344	planthopper, <i>Nilaparvata lugens</i> , exhibiting various responses to density in relation to wing morphism. <i>Entomologia Experimentalis et Applicata, 38</i> , 101–108. doi:10.1111/j.1570-7458.1985.tb03505.x				
342 343 344 345	planthopper, <i>Nilaparvata lugens</i> , exhibiting various responses to density in relation to wing morphism. <i>Entomologia Experimentalis et Applicata, 38</i> , 101– 108. doi:10.1111/j.1570-7458.1985.tb03505.x Jena, M., Pandi, G.G.P., Adak, T., Rath, P.C., Gowda, G.B., Patil, N.B., Golive, P., &				
342 343 344 345 346	 planthopper, <i>Nilaparvata lugens</i>, exhibiting various responses to density in relation to wing morphism. <i>Entomologia Experimentalis et Applicata, 38</i>, 101–108. doi:10.1111/j.1570-7458.1985.tb03505.x Jena, M., Pandi, G.G.P., Adak, T., Rath, P.C., Gowda, G.B., Patil, N.B., Golive, P., & Mahapatra, S.D. (2018). Paradigm shift of insect pests in rice ecosystem and their 				
 342 343 344 345 346 347 	 planthopper, <i>Nilaparvata lugens</i>, exhibiting various responses to density in relation to wing morphism. <i>Entomologia Experimentalis et Applicata, 38</i>, 101–108. doi:10.1111/j.1570-7458.1985.tb03505.x Jena, M., Pandi, G.G.P., Adak, T., Rath, P.C., Gowda, G.B., Patil, N.B., Golive, P., & Mahapatra, S.D. (2018). Paradigm shift of insect pests in rice ecosystem and their management strategy. <i>Oryza, 55</i>, 82-89. doi:10.5958/2249-5266.2018.00010.3 				

351	Khlibsuwan, W., Hanboonsong, Y., Pannangpetch, K., and Sriratanasak W. (2014).
352	Simulation model of brown planthoppers, Nilaparvata lugens stal. (Homoptera:
353	Delphacidae). Khon Kaen Agriculture Journal, 42, 239-248 (In Thai).
354	Kisimoto, R. (1956). Effect of crowding during the larval period on the determination of
355	the wing-form of an adult plant-hopper. Nature, 178, 641–642.
356	doi:10.1038/178641a0.
357	Kikuta, S., Kikawada, T., Hagiwara-Komoda, Y., Nakashima, N., & Noda, H. (2010).
358	Sugar transporter genes of the brown planthopper, Nilaparvata lugens: A
359	facilitated glucose/fructose transporter. Insect Biochemistry and Molecular
360	<i>Biology</i> , 40, 805–813. doi:10.1016/j.ibmb.2010.07.008.
361	Latif, M.A., Omar, M.Y., Tan, S.G., Siraj, S.S., Ali, M.E., & Rafii, M.Y. (2012). Food
362	assimilated by two sympatric populations of the brown planthopper Nilaparvata
363	lugens (Delphacidae) feeding on different host plants contaminates insect DNA
364	detected by RAPD-PCR analysis. Genetics and Molecular Research, 11, 30-41.
365	doi:10.4238/2012.January.9.4.
366	Lin, X., Xu, Y., Jiang, J., Lavine, M, & Lavine, L.C. (2018). Host quality induces
367	phenotypic plasticity in a wing polyphenic insect. Proceedings of the National
368	Academy of Sciences of the United States of America, 115, 7563–7568.
369	doi:10.1073/pnas.172147311
370	Liu, F., Li, X., Zhao, M., Guo, M., Han, K., Dong, X., Zhao, J., Cai, W., Zhang, Q., &
371	Hua, H. (2020). Ultrabithorax is a key regulator for the dimorphism of wings, a

- main cause for the outbreak of planthoppers in rice. *National Science Review*, 7,
- 373 1181–1189. doi.org/10.1093/nsr/nwaa061
- Lu, Z., Yu, X., Heong, K.L., & Hu, C. (2007). Effect of Nitrogen Fertilizer on
- Herbivores and Its Stimulation to Major Insect Pests in Rice. *Rice Science*, *14*,
 56–66. doi:10.1016/S1672-6308(07)60009-2
- Mollah, M.L.R., Samad, M.A., Hossain, M.A., & Khatun, M.F. (2011). Settling and
 feeding responses of brown planthopper to five rice cultivars. *Journal of Crop Production 6*, 10–13.
- 380 Okamura, M., Hashidaa, Y., Hirosea, T., Ohsugia, R., & Aokia, N. (2016). A simple
- method for squeezing juice from rice stems and its use in the high-throughput
 analysis of sugar content in rice stems. *Plant Production Science*, 19, 309–314.
 doi:10.1080/1343943X.2015.1128099
- Rivera, C.T., Ou, S.H., & Iida, T.T. (1966). Grassy stunt disease of rice and its transmission
 by the planthopper *Nilaparvata lugens* Stal. *Plant Disease Reporter*, *50*, 453–456.
 doi:10.48048/tis.2022.5097
- Robbins, N.S., & Pharr, D.M. (1987). Regulation of photosynthetic carbon metabolism
 in cucumber by light intensity and photosynthetic period. *Plant Physiology*, *85*,
 592–597. doi:10.1104/pp.85.2.592
- Romadhon, S., Koesmaryono, Y., & Hidayati, R. (2017). Influence of Climate
- 391 Variability on Brown Planthopper Population Dynamics and Development Time.
- 392 *Vol. 58.* IOP Conference Series: Earth and Environmental Science 58, The 3rd

393	International Seminar On Sciences "Sciences On Precision And Sustainable
394	Agriculture" (ISS-2016) Bogor, Indonesia. doi: 10.1088/1755-1315/58/1/012042
395	Sawada, H., Subroto, S.W.G., Suwardiwijaya, E., Mustaghfirin, & Kusmayadi, A.
396	(1992). Population dynamics of the brown planthopper (Nilaparvata lugens) in
397	the coastal lowland of West Java, Indonesia. The Japan Agricultural Research
398	Quarterly, 26, 88–97.
399	Senthil-Nathan, S.N., Choi, M.Y., Paik, C.H., Seo, H.Y., & Kalavani, K. (2009).
400	Toxicity and physiological effects of neem pesticides applied to rice on the
401	Nilaparvata lugens Stal, the Brown Planthopper. Ecotoxicology and
402	Environmental Safety, 72, 1707-1713. doi:10.1016/j.ecoenv.2009.04.024
403	Saxena, R.C., Okech, S.H., & Liquid, N.S. (1981). Wings morphism in the brown
404	planthopper Nilaparvata lugens. Insect Science and Its Application, 1, 343–348.
405	doi:10.1017/S1742758400000631
406	Simpson, S.J., & Raubenheimer, D. (1993). A multi-level analysis of feeding behavior:
407	the geometry of nutritional decisions. Philosophical Transactions of the Royal
408	<i>Society B</i> , <i>342</i> , 381–402.
409	Smith, C.M., Khan, Z.R., & Pathak, M.D. (1994). Techniques for Evaluating Insect
410	Resistance in Crop Plants. Boca Raton. Florida. USA.
411	Syobu, S., Mikuriya, H., Yamaguchi, J., Matsuzaki, M., & Matsumura, M. (2002).
412	Fluctuations and factors affecting the wing-form ratio of the brown
413	panthopper, Nilaparvata lugens Stal in rice fields. Japanese Journal of Applied
414	Entomology and Zoology, 46, 135-143. doi:10.1303/jjaez.2002.135.

415	Wu, G.R., Yu, X.P., Tao, L.Y., & Ren, Z.J. (1994). Wing dimorphism and migration in
416	the Brown planthopper, Nilaparvata lugens Stal. Series Entomologica, 52, 263–275

- 417 Wu, X., Yu, Y., Baerson, S.R., Song, Y., Liang, G., Ding, C., Niu, J., Pan, Z., & Zeng,
- 418 R. (2017). Interactions between Nitrogen and Silicon in Rice and Their Effects on
- 419 Resistance toward the Brown Planthopper *Nilaparvata lugens*. *Frontiers in Plant*
- 420 *Science*, *8*, 28. doi:10.3389/fpls.2017.00028.
- 421 Xayyasin, T., Khlibsuwan, W., & Tangkawanit, U. (2014). Development of simulation
- 422 model of rice brown planthopper, *Nilaparvata lugens* (Stal.) population. *Khon*423 *Kaen Agriculture Journal*, 42, 453–462. (In Thai).
- 424 Xu, H.J., Xue, J., Bo, L., Zhang, X.C., Zhuo, J.C., He, S.F., Ma, X.F., Jiang, Y.Q., Fan,
- 425 H.W., Xu, J.Y., Ye, Y.X., Pan, P.L., Li, Q., Bao, Y.Y., Nijhout, H.F., & Zhang,
- 426 C.X. (2015). Two insulin receptors determine alternative wing morphs in

427 planthoppers. *Nature*, *519*, 464–467. doi:10.1038/nature14286

- 428 Zhi-Fang, A.N., Ju-Long, Y.U., Juan, P., Chao, Z., & Xiang-Dong, L. (2014).
- 429 Differentiation of wing forms in pure macropterous and brachypterous lineages is
- 430 less subject to photoperiod in rice planthoppers (Hemiptera: Delphacidae). Acta

431 *Entomologica Sinica* 57, 1306–1314. doi:10.3390/genes11010019

- 432 Zhang, C., Mao M.S., & Liu, XD. (2023). Relative contribution of genetic and
- 433 environmental factors to determination of wing morphs of the brown planthopper
- 434 Nilaparvata lugens. Insect Science 30 (1), 208-220. doi: 10.1111/1744-
- 435 7917.13037.



- 446 Figure 1. *Nilaparvata lugens* life stages: (A) eggs; (B) nymph; (C) brachypterous
- 447 form; (D) macropterous form.



449 Figure 2. Sucrose content (mg/gfw) for rice infested with 3 adults of *Nilaparvata*

lugens per clump of rice and for non-infested rice.



455 Figure 3. Rice infestation symptoms at different ages after *Nilaparvata lugens* were

456 released when rice was 40 days old: (A) 68 days old; (B) 75 days old; (C)

457 brachypterous 82 days old; (D) 89 days old; (E) 96 days old; and (F) 96 days old

458 (control rice).

467 SJST MANUSCRIPT TEMPLATE FOR A TABLE FILE

468 Table 1 Food assimilated, honeydew excreted and food ingested by nymph and

469 adult *Nilaparvata lugens* on the Taichung native 1 (TN1) rice variety after 24

470 **hours.**

	Nilaparvata lugens	Food assimilated	Honeydew excreted	Food ingested
	(n=30)	(mg)	(mg)	(mg/day)
		(Mean±SD)	(Mean±SD)	(Mean±SD)
	Nymph	0.72±1.33b*	0.49±0.26b	1.21±1.38b
	Adult	5.75±0.72a	4.85±1.90a	10.60±2.14a
471	*Means followed by	a lowercase letter with	hin the columns are not	significantly different
472	at the 5% level accor	ding to a t-test.		
473				
474				
475				
476				
477				
478				
479				
480				

Rice age	Rice	Population (insects per	Food	demand (mg	/day)
(Days)	stages*	hill	l)			
		Nymph	Adult	Nymph	Adult	Total
40	V	0.00	3.00	0.00	31.80	31.80
47	V	0.00	1.73	0.00	18.35	18.35
54	V	9.69	0.38	11.73	4.08	15.80
61	V	38.46	0.00	46.54	0.00	46.54
68	V	11.77	14.71	14.24	155.94	170.18
75	V	47.81	13.12	57.85	139.02	196.87
82	V	197.02	11.92	238.39	126.38	364.78
89	V	9.54	96.46	11.54	1022.42	1033.9
96	R	2.58	40.95	3.12	434.06	437.17
103	R	0.00	4.10	0.00	43.42	43.42

Table 2 *Nilaparvata lugens* populations and food demands in the rice field experiment for 40- to 103-day-old rice.

483

484 * V = vegetative growth, R= reproductive growth.

487	Table 3 Sugar supply to insect demand ratio and the relative abundance of short-wing

488 form for 40- to	103-day-old rice.
---------------------	-------------------

Rice age (day)	Supply to demand	Adult	Relative	Insect
	(gfw*/insect/hill)	(insect/hill)	abundance	generation
40	0.42	3.00	0.50	
47	0.25	1.73	0.51	
54	0.41	0.38	0.90	1^{st}
61	0.09	0.00**	0.00	
68	0.02	14.71	0.83	
75	0.02	13.12	0.95	2^{nd}
82	0.01	11.92	0.60	
89	0.00	96.46	0.03	
96	0.01	40.95	0.02	3 rd
103	0.04	4.10	0.00	

***gfw = gram fresh weight**

491 ** number of adults in 1^{st} generation died